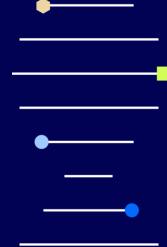
Advancing data standardization via Allotropy

The Allotropy Open-Source Library for Instrument Data Conversion into ASM



Bioanalytical Use Case Enabled by Allotropy Convertor Catalog

Vaccine Bioanalysis: Reimagining E2E workflows to accelerate innovation

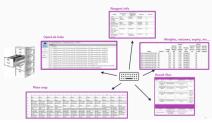
FROM

Sample Management

> Assay Execution



Data Reporting



TO

Automated sample management with an integrated tracking system



Integrated robotics for routine assay execution



Integrated, endto-end data management



Vaccines Bioanalytical Solution

- Vaxbaam is being built using the Benchling Connect platform to enable end-to-end assay data management, data connectivity across upstream and downstream systems. Intent to reuse data to drive future insights.
- Leverage the Allotropy library of ASM converters (~50 in total) in the solution by the end of Q2 2025. Approximately 40% of these are available now or very soon (see below).

✓ Biotek Cytation 7
✓ TapeStation
✓ Biotek Cytation 5
✓ Quant Studio 7 Pro
✓ Quant Studio 5
✓ Bio-Plex 200 Luminex
✓ Nucleocounter NC200
✓ Immunospot Analyzer S6U Universal
✓ FLEXMAP 3D READER
✓ Iris Reader

✓ Sector S 600
✓ Ensight Multimode Plate Reader
✓ Lunatic
✓ QuantStudio 7 Flex
✓ Vi-CELL XR Automated Cell Viability Analyzer
✓ Qiagen
✓ Qiacuity 8, PCR system
✓ Absolute Q
✓ Genesys 150uv scanning
✓ Cellaca MXFL2

Today, you will learn how to leverage & contribute to Allotropy!

Allotropy Overview

Supported ASM Schemas

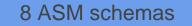
Contributing a Connector



Allotropy is a python library to convert instrument data to ASM



Allotropy coverage is growing, to help drive ASM standardization

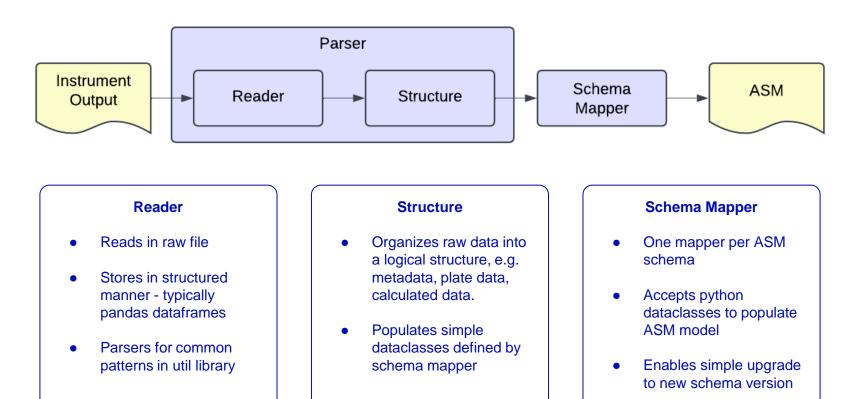


34 parsers

Over 100 supported instruments

ASM schema	Parsers
Cell Counting	6
Electrophoresis	1
Multianalyte Profiling	2
Plate Reader	10
Solution Analyzer	3
Spectrophotometry	7
dPCR	2
qPRC	3

Contributing a connector includes parser & schema mapping



Allotropy has tools to make building connectors to ASM easy

Boilerplate code automation

start a new parser in seconds

File parsing libraries

standard methods for common formats

Auto-generated ASM schema python models

makes populating ASM easy

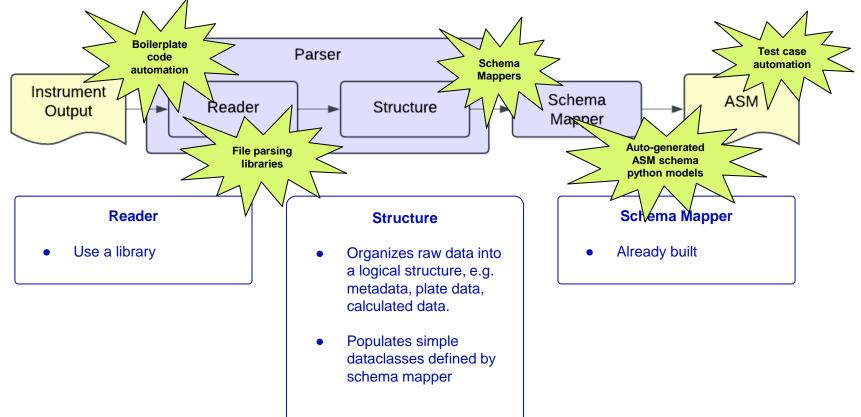
Schema Mappers

makes populating ASM version agnostic

Test case automation

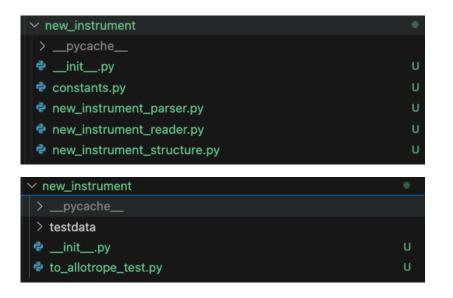
add a test case with no code

Contributing a connector includes parser & schema mapping



Allotropy has a command to create all boilerplate for a new parser

hatch run scripts:create-parser "New Instrument" "2024/06/plate-reader"



from allotropy.allotrope.models.adm.plate_reader.rec._2024._06.plate_reader import (Model,

Mapper,

from allotropy.named_file_contents import NamedFileContents
from allotropy.parsers.new_instrument.constants import DISPLAY_NAME
from allotropy.parsers.new_instrument.new_instrument_reader import (
 NewInstrumentReader,

from allotropy.parsers.new_instrument.new_instrument_structure import (
 create_measurement_groups,
 create_metadata,

from allotropy.parsers.release_state import ReleaseState from allotropy.parsers.utils.pandas import map_rows from allotropy.parsers.vendor_parser import VendorParser

class NewInstrumentParser(VendorParser[Data, Model]): DISPLAY_NAME = DISPLAY_NAME RELEASE_STATE = ReleaseState.WORKING_DRAFT SUPPORTED_EXTENSIONS = NewInstrumentReader.SUPPORTED_EXTENSIONS SCHEMA_MAPPER = Mapper

Allotropy has a growing library of utilities for parsing input files

Section reader

##8	LOCK	(S=	12	

##DLL	CK2= 1																			
	BNC													nce						
	empera	ture(iC) A1									A10						B4		Bŧ
6	1.3	28 (0.387		422		446		721	-0.	22		902	0.1	81	0.9	43	1.3	87	-6
e		21525	1.2	21775	0.6	0375	0.8	9175	0.5	7575	1.3	6875	0.2	8075	1.6	0475	2.3	7175	2.1	23
e		94162!		. 3966	25		7596	25		1456	25	-0.	8806	25		4616	25	0.5	0137	5
			A3 A4						A10						В4		B6		B8	B
	4.5	81298	5556530	9714	0.7	7015	3899	3601	9363	-0.	4858	3478	5000	4584	6	0.5	5957	0352	6894	11
~End																				
Group	: Posi	tiveC	ontrol																	
Sampl	le Wel		Concent	trati	on	Val	ues	Mea	nVal	ue	Std	.Dev		CV%	Wel	lPla	teNa	me		
CONTR	IOL_748	01 <i>i</i>	A1	4.5	01	1.0	64	1.5	34	144	.129	BNC	1_69	9835	42_9	6w				
		0.77	3			BNC	H_69	9835	42_9	6w										
E		-0.1	30			BNC	H_69	9835	42_9	6w										
E		0.74				BNC	H_69	9835	42_9	6w										
		1.25				BNC	H_69	9835	42_9	6w										
		-0.0	55			BNC	H_69	9835	42_9	6w										
		1.58				BNC	н_69	9835	42_9	6w										
		-0.1	54			BNC	н_69	9835	42_9	6w										
Group	Colum		Formula	a Nam	e	For	mula	Pre	cisi	on	Nota	atio								
	iample	!Samp	pleName	es	3 d	ecim	alp	lace		Num	eric									
2 V	/ell	!Wel	lIDs	3 d	ecim	alp	lace		Num	eric										
3 (oncent	ratio	n !Sa	ample	Desc	ript	or	3 d	ecim	alp	lace	s	Num	eric						
4 \	alues	!Wel	lValue	s 3 d	ecim	alp	lace		Num	eric										
5 N	leanVal	ue /	Average	e(!We	llVa	lues		3 d	ecim	alp	lace		Num	eric						
6 5	itd.Dev		Stdev(Well	Valu	es)	3 d	ecim	alp	lace		Num	eric							
	.V% Cv(Well!	/alues) 3 d	ecim	alp	lace		Num	eric										
о I.	(a) 101 a	holdon.	- 114	1101	-		3.4		-1 -	1		Martin								

<pre>sub_reader in BlockListiter_blocks_reader(reader);</pre>
<pre>if sub_reader.match("^Group"):</pre>
if "WellPlateName" in assert_not_none(
<pre>sub_reader.get_line(sub_reader.current_line + 1), msg="Unable to get columns from group block",</pre>
<pre>group_blocks.append(GroupBlock.create(sub_reader)) elif sub reader.match("^Plate");</pre>
header_series = PlateBlock.read_header(sub_reader)
<pre>plate block cls = PlateBlock.get plate block cls(header series)</pre>
header = plate_block_cls.parse_header(header_series)
<pre>export_format_to_data_format = {</pre>
ExportFormat.TIME_FORMAT.value: TimeData,
ExportFormat.PLATE_FORMAT.value: PlateData,
<pre>data_format: type[TimeData] type[PlateData] = get_key_or_error("export format", header.export_format, export_format_to_data_format</pre>
<pre>block_data = data_format.create(sub_reader, header)</pre>
<pre>plate_blocks[header.name] = plate_block_cls(</pre>
header=header,
block_data=block_data,
elif not sub_reader.match(""Note"):
<pre>msg = f"Expected block '{sub_reader.get()}' to start with Group, Plate o</pre>

Pandas row accessors

leasurement(

measurement_identifier=random_uuid_str[]], timestamp=data[str, "Analysis date/time"], sample_identifier=data[str, "Sample ID"], cell_type_processing_method=data.get(float, "Minimum Diameter (µm)"), minimum_cell_diameter_setting=data.get(float, "Maximum Diameter (µm)"), cell_density_dilution_factor=data.get(float, "Maximum Diameter (µm)"), cell_density_dilution_factor=data.get(float, "Dilution"), viability=data[float, "Viability (%)"], viabile_cell_density=data[float, "Viable (x10^6) cells/mL"], total_cell_density=data.get(float, "Average diameter (µm)"), average_total_cell_diameter=data.get(float, "Average viable diameter (µm)"), viable_cell_count=viable_cell_count,

average_total_cell_circularity=data.get(float, "Average circularity"),
average_viable_cell_circularity=data.get(
 float, "Average viable circularity"



analyst=get_val_from_xml(environment, "Experimenter"),|
analytical_method_identifier=get_val_from_xml_or_none(
 file_information, "Assay"

data_system_instance_identifier=get_val_from_xml(environment, "Computer"),

2

Allotropy has a tool to generate python models for ASM schemas

@dataclass(kw_only=True)

class PlateReaderDocumentItem:

measurement_aggregate_document: MeasurementAggregateDocument analyst: TStringValue | None = None electronic_project_record: ElectronicProjectRecord | None = None submitter: TStringValue | None = None

@dataclass(kw_only=True)

class PlateReaderAggregateDocument: plate_reader_document: list[PlateReaderDocumentItem] analysis_sequence_document: AnalysisSequenceDocument | None = None calculated_data_aggregate_document: CalculatedDataAggregateDocument | None = None custom_information_aggregate_document: CustomInformationAggregateDocument | None = (None

data_system_document: DataSystemDocument | None = None device_system_document: DeviceSystemDocument | None = None electronic_project_record: ElectronicProjectRecord | None = None electronic_signature_aggregate_document: ElectronicSignatureAggregateDocument | None = { None

processed_data_aggregate_document: ProcessedDataAggregateDocument | None = None
statistics_aggregate_document: StatisticsAggregateDocument | None = None

@dataclass(kw_only=True)

class Model:

field_asm_manifest: AdmCoreREC202409ManifestSchema | str
plate_reader_aggregate_document: PlateReaderAggregateDocument | None = None

@dataclass(kw_only=True)

detection_type: TStringValue | None = None electronic_project_record: ElectronicProjectRecord | None = None error_aggregate_document: ErrorAggregateDocument | None = None image_aggregate_document: ImageAggregateDocument | None = None measurement_time: TDateTimeStampValue | None = None processed_data_aggregate_document: ProcessedDataAggregateDocument | None = None statistics_aggregate_document: StatisticsAggregateDocument | None = None compartment_temperature: TQuantityValueDegreeCelsius | None = None mass concentration: TOuantityValuePicogramPerMilliliter | None = None electropherogram_data_cube: TDatacube | None = None chromatogram data cube: TDatacube | None = None processed data document: ProcessedDataDocument | None = None absorption_profile_data_cube: TDatacube | None = None fluorescence: TQuantityValueRelativeFluorescenceUnit | None = None fluorescence emission profile data cube: TDatacube | None = None luminescence: TQuantityValueCountsPerSecond | TQuantityValueRelativeLightUnit | None = (

luminescence_profile_data_cube: TDatacube | None = None

hatch run scripts:download-schema <url>

hatch run scripts:generate-schemas

Allotropy has a schema mappers to make populating ASM easy

@dataclass(frozen=True)

class Measurement:

Measurement metadata
type_: MeasurementType
device_type: str
identifier: str
sample_identifier: str
location_identifier: str

Optional metadata

well_plate_identifier: str | None = None
detection_type: str | None = None
sample_role_type: SampleRoleType | None = None

Measurements

absorbance: float | None = None
fluorescence: float | None = None
luminescence: float | None = None

Settings

detector_wavelength_setting: float | None = None detector_bandwidth_setting: float | None = None excitation_wavelength_setting: float | None = None excitation_bandwidth_setting: float | None = None wavelength_filter_cutoff_setting: float | None = None detector_distance_setting: float | None = None scan_position_setting: ScanPositionSettingPlateReader | None = None detector_gain_setting: str | None = None detector_carriage_speed: str | None = None compartment_temperature: float | None = None number_of_averages: float | None = None Removes need for dev to create nested document

One schema mapper per ASM schema - used by all parsers

Enables upgrading schema version without changing parsers

get_ultraviolet_absorbance_measurement_document(
self, measurement: Measurement
> MeasurementDocument:
return MeasurementDocument(
<pre>measurement_identifier=measurement.identifier,</pre>
<pre>sample_document=selfget_sample_document(measurement),</pre>
<pre>device_control aggregate_document=DeviceControlAggregateDocument(</pre>
<pre>device_control_document=[</pre>
DeviceControlDocumentItem(
<pre>device_type=measurement.device_type,</pre>
<pre>detection_type=measurement.detection_type,</pre>
<pre>detector_wavelength_setting=quantity_or_none(</pre>
TQuantityValueNanometer,
<pre>measurement.detector_wavelength_setting,</pre>
<pre>number_of_averages=quantity_or_none(</pre>
TQuantityValueNumber, measurement.number_of_averages
<pre>detector_carriage_speed_setting=measurement.detector_carriage_speed,</pre>
<pre>detector_gain_setting=measurement.detector_gain_setting,</pre>
<pre>detector_distance_settingplate_reader_=quantity_or_none(</pre>
TQuantityValueMillimeter,
<pre>measurement.detector_distance_setting,</pre>

8

Allotropy has a testing library that allows no-code test addition

hatch run test tests/parsers/moldev_softmax_pro/ --overwrite

hatch run test tests/parsers/moldev_softmax_pro/ --filter <test_case>

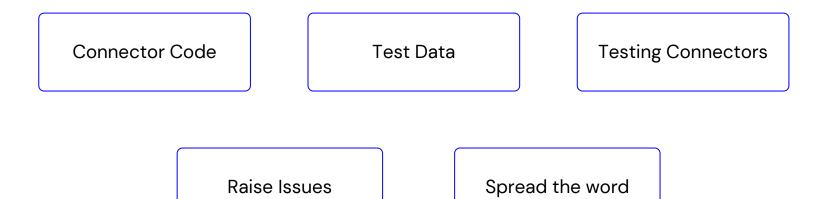
Automatically detects test cases in testdata/

Validates output against ASM schema Overwrite expected output easily while iterating

~100 lines of code of non-boilerplate code to add a simple parser!

1	import pandas as pd	24 🗸	def create_measurement_group(data: SeriesData) -> MeasurementGroup:
2		25	# This function will be called for every row in the dataset, use it to create
3	<pre>from allotropy.named_file_contents import NamedFileContents</pre>	26	# a corresponding measurement group.
4	<pre>from allotropy.parsers.lines_reader import determine_encoding</pre>	27	W 6-13 seconds are assessed in a 232-bit show an add in a 233-bit show at a 23-bit
5	from allotropy.parsers.utils.pandas import (28 29	# Cell counts are measured in cells/mL, but reported in millions of cells/mL viable_cell_density = float(
6	df_to_series_data,	30	Decimal(data[float, "Live Cells/mL"]) / Decimal("1000000")
7	read_csv,	31	
		32	total_cell_density = data.get(float, "Total Cells/mL")
8	read_excel,	33	<pre>if total_cell_density:</pre>
9		34	<pre>total_cell_density = float(Decimal(total_cell_density) / Decimal("1000000"))</pre>
10		35	<pre>dead_cell_density = data.get(float, "Dead Cells/mL")</pre>
11		36	<pre>if dead_cell_density:</pre>
12 🗸	class RevvityMatrixReader:	37	<pre>dead_cell_density = float(Decimal(dead_cell_density) / Decimal("1000000"))</pre>
13	SUPPORTED_EXTENSIONS = "csv,xlsx"	38	
14	data: pd.DataFrame	39 40	errors = data.get(str, "Errors:", validate=SeriesData.NOT_NAN) return MeasurementGroup(
15		40	measurements=[
16 ~	<pre>definit(self, named_file_contents: NamedFileContents) -> None:</pre>	42	Measurement (
10 1	if named_file_contents.extension == "csv":	43	<pre>measurement_identifier=random_uuid_str(),</pre>
		44	# NOTE: instrument file does not provide a timestamp, but it is required by ASM, so pass
18	<pre>contents = named_file_contents.contents.read()</pre>	45	# EPOCH to signal no timestamp.
19	encoding = (46	<pre>timestamp=DEFAULT_EPOCH_TIMESTAMP,</pre>
20	<pre>determine_encoding(contents, named_file_contents.encoding)</pre>	47	<pre>sample_identifier=data[str, "Well Name"],</pre>
21	<pre>if isinstance(contents, bytes)</pre>	48	viability=data[float, "Viability"],
22	else None	49 50	<pre>total_cell_count=data.get(float, "Total Count"), total_cell_density=total_cell_density,</pre>
23)	50	total_cell_oensity=total_cell_density, average_total_cell_diameter=data.get(float, "Total Mean Size"),
24	<pre>named_file_contents.contents.seek(0)</pre>	52	viable_cell_count=data.get(float, "Live Count"),
25	<pre>df = read_csv(named_file_contents.contents, encoding=encoding)</pre>	53	viable_cell_density=viable_cell_density,
26	else:	54	average_live_cell_diameter=data.get(float, "Live Mean Size"),
27	<pre>df = read_excel(named_file_contents.contents)</pre>	55	<pre>dead_cell_count=data.get(float, "Dead Count"),</pre>
28	<pre># Reading a percent value (50%) in read_excel results in a decimal: 0.5</pre>	56	<pre>dead_cell_density=dead_cell_density,</pre>
29	# Detect and adjust value back to 0-100%	57	<pre>average_dead_cell_diameter=data.get(float, "Dead Mean Size"),</pre>
		58	errors=[
30	<pre>first_row = df_to_series_data(df, 0)</pre>	59 60	Error(error=error) for error in (errors.split(",") if errors else [])
31	<pre>viability = first_row[str, "Viability"]</pre>	61],
32	<pre>if "%" not in first_row[str, "Viability"] and float(viability) < 1:</pre>	62	
33	df["Viability"] = df["Viability"] * 100	63	
34	self.data = df	64	

There are 5 easy ways to contribute



Thank you!

Questions?